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**Efficacy of *Zonocercus Variiegatus* (Grasshopper) Meal on *Clarias Gariiepinus* Juveniles – Growth Rate in Lafia, Nasarawa State, Nigeria.**

Ayuba Scholastica Onyaweyo<sup>1</sup>, Abdulrahman Itopa Suleiman<sup>2\*</sup>, Hamza Magaji<sup>3</sup>, Nana-mariam Abdullahi<sup>4</sup>, Ahmad Abdulrazaq Itopa<sup>2</sup>, Muhammad Bilal<sup>5</sup>, and Ummulhanni Oyiza Obansa<sup>6</sup>

Department of Zoology, Federal University of Lafia, Lafia, Nasarawa State, Nigeria<sup>1</sup>, Department of Science Laboratory Technology, Kogi State Polytechnic, Lokoja, Nigeria<sup>2</sup>, Department of Biological Sciences, Federal University of Technology, Babura, Jigawa State, Nigeria<sup>3</sup>, Department of Biological Sciences, Confluence University of Science and Technology, Osara, Kogi State, Nigeria<sup>4</sup>, Ministry of Agriculture and Natural Resources, Bauchi State, Nigeria<sup>5</sup>, Faculty of Veterinary Medicine, University of Maiduguri, Borno State, Nigeria<sup>6</sup>.

Author for Correspondence: itopa2020@gmail.com/scholasticaayuba@gmail.com/https://dx.doi.org/10.4314/sokjmls.v9i3.14

**Abstract**

The residents of the north-eastern part of Nigeria like eating edible grasshoppers, which have an amino acid profile that is comparable to fish meal. In terms of amino acid composition, these grasshoppers are just as rich as fish meal. This study investigated the effects of feeding juveniles *Clarias gariiepinus* different amounts of Grasshopper meal (GM) in their diet. Twenty-four (24) *Clarias gariiepinus* juveniles were bought, acclimated for seven days and divided into five treatments groups (Control, T1, T2, T3, and T4). The grasshopper was ground into powder form after being oven-dried to a uniform weight. Five diets were created, each with a different inclusion level of 0%, 30%, 40%, 50% and 60 % respectively. The proximate analysis showed that GM contained 60.90% Crude protein, 12.0% Ether extract, 1.0% Ash content and 17.0% crude fibre accordingly. Diet T3 had the lowest survival rate at 60%, while diets T1, T4, TC and T2 all had survival rate of 75%. However, there was significant difference ( $P < 0.05$ ) between the two diets. In all of the treatments that were looked at, the feed conversion ratio (FCR) revealed significant variations ( $p < 0.05$ ). Increase mean Length (IML) varied significantly ( $p < 0.05$ ) with T4 gaining the most length (12.67cm) and T3 gaining the least (5.58cm). With the exception of T2 and T3, the physico-chemical parameters of water in the experimental tank were within the acceptable range for *C. gariiepinus* optimum growth. It is recommended that Grasshopper meal (GM) be added to the feed of *C. gariiepinus* for optimum growth performance and lower production costs.

**Keywords:** *Clarias gariiepinus*, Feed conversion ratio (FCR), Grasshopper meal (GM), Increase mean length (IML), Juveniles

**Introduction**

Protein, an essential macronutrient that performs numerous bodily activities is primarily found in fish. It contains omega-3 fatty acids as well as vitamin D and B2 (riboflavin). Protein is typically composed of carbon (50%), nitrogen (16%), oxygen (21.5%) and hydrogen (6.5%), with certain protein also containing small amount of sulfur (2.12%) (Halver and Hardy, 2002). One of the distinguishing species in the subclass *Actinopterygii*, order *Siluriformes*, and family *Clariidae* is the African catfish, *Clarias gariiepinus*. The habitat needs of the African catfish species are quite common and pervasive. Although, it is predatory and omnivorous, it has a relatively high dietary protein requirement of between 40 and 50 percent of crude protein on a daily weight basis (Balogun and Dabrowski, 1992). There is a rising demand for seafood as a result of the human rapid growth in human population and rising level of life (Daniel, 2018; Stankus, 2021). Fish feed is presently very expensive both imported and locally produced ones (Falaye, 2003). The use of other sustainable sources as alternatives for fish meal in relation to the production of formulated feed has reduced the dependence on fish meal. The rise in global population has increased the demand for protein source, which has inevitably impacted the aquaculture industry to produce high yielding fish with lower cost (Barroso *et al.*, 2014; Henry *et al.*, 2015). The global aquaculture production of farmed food fish in 2018 reached 82 million tons

with a value of USD 400 billion, representing around half of the total world fish production for human consumption (FAO, 2020).

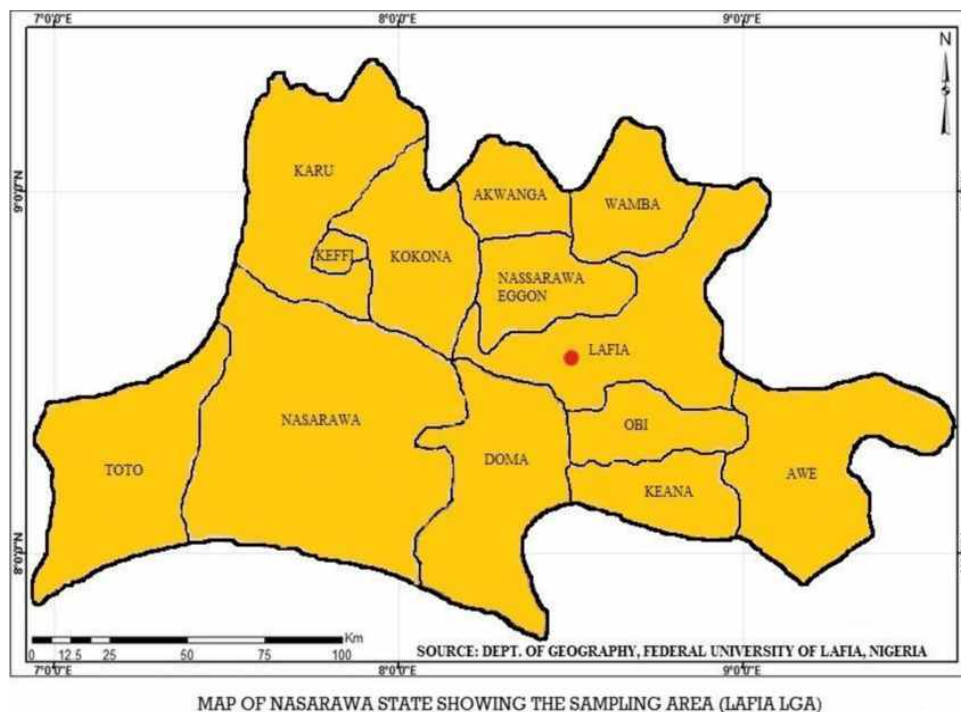
Insects have gained a lot of interest recently as a source of protein for both humans and livestock. Insects are capable of rapid growth and reproduction, high feed conversion efficiency, and bio-waste rearing (Van Huis *et al.*, 2013; Makkar *et al.*, 2014). An average of 2 kg of feed biomass may be converted into 1 kg of insect biomass (Collavo *et al.*, 2005). Insects are one of the sustainable and promising alternatives to fishmeal for use in aqua diets (Henry *et al.*, 2015). The utilization of insect meals as a substitute for fish meal has been successfully evaluated for different number of fish species such as the European sea bass (Gasco *et al.*, 2016) trout *Oncorhynchus mykiss* (Walbaum) (Renna *et al.*, 2017); Nile tilapia; *Oreochromis niloticus* by (Sanchez muros *et al.*, 2016) gilthead seabream (Karapanagiotidis *et al.*, 2014); the Siberian sturgeon, *Acipenser baerii* Brandt (Zarantoniello *et al.*, 2021); the clownfish, *Amphiprion ocellaris* Cuvier (Vargas-Abúndez, *et al.*, 2019) or the zebrafish, *Danio rerio* (Hamilton) (Zarantoniello *et al.*, 2019) and shrimps (Motte *et al.*, 2019). There are several reports on the use of some alternative animal protein feedstuffs to fish

meal such as: Black soldier fly larvae with about 40-44% crude protein (Makkar *et al.*, 2014; Tran *et al.*, 2015), Locust with high level of protein, minerals and vitamins, and contain 50-65% crude protein (Makkar *et al.*, 2014), Sliver cyprinid (*Rastrioneobola argentea*) with protein values range from 19.1-21.8% (Bille and Shemkai, 2006). The similarity in the quality of the amino acid profiles of fish and grasshopper meal encourages the conclusion that these grasshoppers are as rich in amino acids as fishmeal. When they fall into a fish pond, according to Madu *et al.*, 2003, fish eat them while they are still alive. Therefore, it is important that insects be used as the primary protein source to increase the growth rate of juvenile *Clarias gariepinus*.

## 2.0 Materials and methods

### 2.1 Study site

The research was conducted in Lafia, Nasarawa State at the College of Agriculture, Science and Technology Experimental farm of the Department of Fisheries Technology, Lafia, Nasarawa State. Lafia is a town in North Central Nigeria; it is the capital of Nasarawa State with its current metro area population of 361,000 by 2022 which is about 3.74% increase from 2021 (NIMET, 2021).



**Figure 1: Map of Nasarawa state showing the study site**

## 2.2 Grasshopper collection and processing

A sample of a grasshopper that are edible was purchased from a market in Maiduguri, Borno state, Nigeria. Prior to proximate analysis, the sample was maintained in a cold chamber after being oven dried at 55°C for 24 hours to generate grasshopper meal, which was then crushed to a fine powder using a mortar and pestle. The moisture content, total ash, crude fat, crude fiber, and crude protein of the target insect powder were then calculated utilizing control laboratory techniques (AOAC, 2005).

## 2.3 Preparation of the experimental diet

The food composition and concentration for each treatment, which was carried out using the technique outlined by Olurin *et al.* (2006), are shown on Tables 1. Fishmeal (FM), maize meal, soybeans, vitamin, bone meal, cassava starch, salt mineral, and dicalcium phosphate (DCP) were all purchased as raw materials from a nearby livestock feed plant. The feedstuffs received the necessary processing, were individually ground using a home hammer milling machine, packaged, and kept for use. Grasshopper meal was thoroughly combined with cassava starch as a binder and added at 0g (control), 30g, 40g, 50g, and 60g. Using a local small pelleting machine (KCM, Y132m-4), the resulting dough was formed into pellets with a 2mm diameter. The wet pellet was air-dried for 24 hours in the shade before being stored for later usage in the laboratory in a new airtight polythene bag.

## 2.4 Experimental design

The experiment was created using a modified version of the methodology of Akinrotimi *et al.* (2011). One hundred and twenty (120) *Clarias gariepinus* juveniles were obtained from the College of Agriculture, a recognized fish farm in

Lafia, acclimated for seven days and then randomly placed in a concrete tank that measured 148 cm in length and 81 cm in width. Each tank received 24 fish, representing the five treatments of 0g (control), 30g, 40g, 50g, and 60g, which were referred to as T1, T2, T3, and T4 correspondingly. Weekly fish weigh-ins were used to modify the feeding rate. The investigation lasted for eight weeks. Fish growth response to diet was assessed using weekly weight of fish and feed provided. The system's water quality was observed and maintained within acceptable bounds at that time.

## 2.5 Statistical analysis

( $P < 0.05$ ) was regarded as statistically significant, and one-way analysis of variance (ANOVA) was performed on all data using SPSS version 21.0 (SPSS Inc., Chicago IL, USA) to determine the specific growth rate (SGR), feed conversion ratio (FCR), percentage mean weight gain (PMWG), percentage survival rate (PSR), and survival rate (SR) across treatment groups.

## 3.0 Results

### 3.1 Experimental Diet composition and Proximate analysis of Grasshopper meal

All treatment groups received the following percentages of the experimental diet: fish meal 32%, soya beans 33%, maize 30%, vitamin premixes 1%, bone meal 0.5%, lysine 0.5%, methionine 0.5%, palm oil 1%, cassava starch 1%, and salt 0.5%. Grasshopper meal was added at various inclusion levels of 30%, 40%, 50%, and 60% (Table 1). According to the proximate composition (Table 2), grasshopper meal contains 4.00% dry matter, 5.1% moisture, 60.90% crude protein, 12.0% ether extract, 1.0% ash, and 17.0% crude fiber.

**Table 1: The Composition of the Experimental diet**

<b>Ingredient</b>	<b>Composition (%dry matter)</b>	<b>Control</b>	<b>T<sub>1</sub>(30%)</b>	<b>T<sub>2</sub>(40%)</b>	<b>T<sub>3</sub>(50%)</b>	<b>T<sub>4</sub>(60%)</b>
Fish meal	32.00	32.00	31.7	31.6	31.5	31.4
<b>Grasshopper</b>	64.00	0	0.3	0.4	0.5	0.6
Soya bean	33.00	33.0	33.0	33.0	33.0	33.0
Maize	30.00	30.0	30.0	30.0	30.0	30.0
Vitamin premixes	1.0	1.0	1.0	1.0	1.0	1.0
Bone meal	0.5	0.5	0.5	0.5	0.5	0.5
Lysine	0.5	0.5	0.5	0.5	0.5	0.5
Methionine	0.5	0.5	0.5	0.5	0.5	0.5
Palm oil	1.0	1.0	1.0	1.0	1.0	1.0
Cassava starch	1.0	1.0	1.0	1.0	1.0	1.0
Common salt	0.5	0.5	0.5	0.5	0.5	0.5
<b>Total</b>	<b>100.0</b>					

**Table 2: Proximate composition of Grasshopper meal**

<b>Sample</b>	<b>%Dry matter</b>	<b>%Moisture content</b>	<b>%Crude Protein</b>	<b>%ether extract</b>	<b>%Ash content</b>	<b>%Crude fibre</b>
Grasshopper meal	4.00	5.1	60.90	12.0	1.0	17.0

**3.2 Feed Conversion Ratio (FCR)**

T3 had the highest Feed Conversion Ratio (68.71), followed by T2 (32.44), Control (18.76), and T4 (14.51), while T1 had the lowest Feed Conversion Ratio (11.86). The Feed Conversion Ratio between T1, T2, and T3, however, differed significantly (P 0.05) from T1, T2, and T3. Control and T4 don't, however, display any statistical significance (P>0.05).

The specific growth rate (SGR) was highest in T1 (37.35) and lowest in T3 (6.47), followed by T4 (30.54), Control (23.62), T2 (13.66) and T3. However, there was a significant difference (P0.05) between T2, T3, and Control but not

between T1 and T4, T1, T2, or T4.

The survival rate in T1 was high at 87.5%, with a significant difference (P0.05) between T1, T3, and T4. However, there was no change between control and T2 that was statistically significant (P 0.05). PSR, or Percentage Survival Rate. The fish fed with T1 had the best survival rate (87.5%), while the fish fed with T2 and T3 had the worst survival rate (66.66%). With the exception of T2 and T3, where there was a significant difference (P0.005), none of the experimental diets had an impact on the fish's survival rate (P>0.05) (Table 3)

**Table 3: Feed conversion ratio (FCR) and other parameters:**

Parameters	Feed Conversion ratio (FCR)	Specific growth rate (SGR)	Survival rate (SR)	Percentage survival rate (PSR)
Control	18.76 <sup>c</sup>	23.62 <sup>b</sup>	16 <sup>c</sup>	66.66 <sup>c</sup>
T1	11.86 <sup>d</sup>	37.35 <sup>a</sup>	21 <sup>a</sup>	87.5 <sup>a</sup>
T2	32.44 <sup>b</sup>	13.66 <sup>c</sup>	16 <sup>c</sup>	66.66 <sup>c</sup>
T3	68.71 <sup>a</sup>	6.47 <sup>d</sup>	15 <sup>d</sup>	62.5 <sup>c</sup>
T4	14.51 <sup>c</sup>	30.54 <sup>a</sup>	17 <sup>b</sup>	70.83 <sup>b</sup>

\*SGR- Specific Growth rate, PMWG- Percentage mean weight gain, FCR- Feed Conversion Ratio and PSR- percentage Survival Rate.

\*Mean values with the same superscript along rows are not significantly different (p>0.005)

\*Mean Values with Different superscripts along rows are significantly different (P<0.005).

**3.3 Growth performance of *Clarias gariepinus* Juveniles**

After 62 days, the results of *Clarias gariepinus* given the experimental diet were displayed in Table 4. Before the trial, the initial weights of every fish given with every experimental diet were the same. After 62 days of the experiment, fish with a 30% inclusion had the highest mean weight gain (23.25g) and weight gain (27.25g). However, T4 had the largest increase in length, followed by T1, Control, T2, and T3, which had

the smallest increase at 5.58 cm. Nevertheless, there was a significant difference in the lengthening of T1 between the experimental group and the control group. The experimental fish were fed diets containing various amounts of grasshopper meal, and weight gain revealed that T1 had the fastest growth trend, followed by T4, then TC. T2 and T3 showed sluggish growth. The experimental fish's growth rate varied significantly (P0.005) between T1, Control, and T4 (Figure 1).

**Table 4: Morphometric indices of the fishes at different treatment groups.**

Parameters	Number of fish stocked	Weight gained				Length gained		
		Initial Weight	Final Weight	Mean Weight Gained	% Mean Weight Gained	Initial Length	Final Length	Increased Mean Length (IML)
Control	24	4.09	18.70 <sup>b</sup>	14.65 <sup>c</sup>	358.19 <sup>c</sup>	6.46	15.36 <sup>b</sup>	8.90 <sup>a</sup>
T1	24	4.00	27.25 <sup>a</sup>	23.16 <sup>a</sup>	566.25 <sup>a</sup>	6.72	18.30 <sup>a</sup>	11.28 <sup>c</sup>
T2	24	4.10	12.56 <sup>c</sup>	8.47 <sup>d</sup>	207.09 <sup>c</sup>	6.80	15.26 <sup>b</sup>	8.46 <sup>d</sup>
T3	24	4.00	8.09 <sup>d</sup>	4.00 <sup>d</sup>	97.79 <sup>d</sup>	6.72	12.3 <sup>c</sup>	5.58 <sup>d</sup>
T4	24	4.08	23.03 <sup>a</sup>	18.94 <sup>b</sup>	463.08 <sup>b</sup>	6.66	19.33 <sup>a</sup>	12.67 <sup>b</sup>
<b>Total</b>	<b>120</b>							

\*MWG-Mean Weight gain, IML- Increase in mean length.

\*Mean values with the same superscript along rows are not significantly different (p>0.005)

\*Mean Values with Different superscripts along rows are significantly different (P<0.005)

**Table 5: Water Quality Parameter Range During the Experimental Period**

Parameter	Control	T1 (30%)	T2 (40%)	T3 (50%)	T4 (60%)	Normal Range for fish culture
	Min -Max	Min -Max	Min -Max	Min -Max	Min-Max	
Temperature	24 <sup>0</sup> C -26 <sup>0</sup> C	23 <sup>0</sup> C -29 <sup>0</sup> C	22 <sup>0</sup> C -25 <sup>0</sup> C	23 <sup>0</sup> C -26 <sup>0</sup> C	24 <sup>0</sup> C- 28 <sup>0</sup> C	25 <sup>0</sup> C -31 <sup>0</sup> C
pH	6.5-7.0	7.0-8.0	5.5 -6.8	6.0 -7.5	6.5 -7.5	6.5 -8.5
D.P (mg/l)	6.8 -6.9	5.0 – 9.5	4.0 -4.5	3.2 -4.8	3.2 – 7.8	5mg/l

#### 4.0 Discussion

One cannot overstate the importance of fish eating as a source of protein for people. The impact of grasshopper feed on the juvenile *C. gariepinus* growth rate was examined in this study. According to the results of the nutritional analysis, grasshopper meal has a high crude protein content of 60.90%. This value is extremely high and could entirely replace fishmeal in fish diet. The amount is consistent with the findings of Njidda *et al.* (2010) who found that the diet of *C. gariepinus* fingerlings contained 64.32% crude protein. Contrarily, Subhachae *et al.* (2010) report stated that the crude protein level was 18.39%. This figure is also comparable to fishmeal made from clupeids by Okoye *et al.* (2003) which had a crude protein content of 68.47%. Crude protein, ether extract, ash content, and crude fiber are all present in grasshopper meal in that order. The importance of these components makes it an excellent nutritional feed for the manufacturing of fish feed.

The feed conversion ratio (FCR) for T1 was high at 11.86, which is consistent with the findings of Haruna, 2003, who found a 2.32 FCR with 20% inclusion of grasshopper meal. This might be due to the experiment's high grasshopper meal inclusion level, 100% feed conversion, and therefore the animals' quick growth. Similar to this, Adikwu (2003) noted that fish use feed more effectively the lower the FCR. According to Finke, 2015 research, T1 had the highest specific growth rate (SGR), which is in contrast to Okoye *et al.* 2004 findings that claimed 10% grasshopper meal produced higher growth results. This may be due to protein utilization as the protein level was 30% more appropriate than the other treatments. Fishes

require specific amounts of protein, and once the body has met those needs, the remaining protein is wasted. According to Omeru and Solomon (2014), the greater the SGR value, the higher the feed quality. T1 had a high percentage survival rate (PSR) of 87.5%. This is consistent with the results of Ogunji *et al.* (2007), who observed a comparable percentage survival rate of 73-80% in tilapia *O. niloticus* fingerlings fed on diets made with inclusion levels of *M. domestica* maggot meal of 45% and 68%. This can be because the water quality is good and the inclusion level is 30%, which is optimal for growth. The percentage mean weight gain (PMWG) in T1 was high at 566.25%, which is consistent with Abanikannda (2012) findings of 801.35%.

The findings of Gbadamosi *et al.* (2007), who indicated that 30% grasshopper feed exhibited decreased weight gain in *C. gariepinus*, are in contradiction. The outcome of the current study could be attributed to the protein content and feed's quality as well as the fact that grasshoppers contain a growth-promoting metabolite that enhances the function of the intestinal flora, improving digestion and enhancing energy utilization, which in turn led to better fish growth. (Abdel-Tawwab *et al.* 2010).

T4 had the largest length, with 12.67 cm, according to the Increased Mean Length (IML). This result contrasts with those of Okoye *et al.* (2004) and is in agreement with Madu *et al.* (2003) which found an increase in the mean length of Tilapia fish to 8.65 cm. This might be a result of the low water quality, which impairs their food intake and causes the fish to become

emaciated and grow longer rather than gaining weight. The surviving fish would degrade the water quality since they would only consume what they needed for protein. It would slow their growth, prevent them from gaining weight, and lengthen them. Factors as a result of high ammonia, high hydrogen sulphide and high carbon dioxide in the water. Except for T2 and T3, which exhibited a drop in pH (5.5, 5.0), temperature (22°C, 23°C), and D.O. (4.0, 3.2).

The physico-chemical parameters seen in the various treatment groups were within the permissible range for fish growing. This could be explained by the delayed development being caused by low feed use. Dissolved oxygen, pH, and temperature are some of the most significant physico-chemical factors that might affect fish growth (Chukwuma and Henry, 2012). Since dissolved oxygen has an impact on fish and the growth of other organism, survival, distribution, behavior, physiology, oxygen depletion in water can result in poor fish nutrition, hunger, stunted growth, and increased fish mortality either directly or indirectly (Bhatnager, 2000). When the pH falls below 4, catfish will perish from the acidity of the water. An acceptable pH value is between (6.5 and 7.5).

#### 4.1 Conclusion

The experimental fish in this study avidly ingested grasshopper meal, which significantly accelerated their growth. The T1 with 30% inclusion and T4 with 60% inclusion groups showed the best overall results. The amount of crude protein in grasshopper meal is high (60.90%), and it also contains other nutrients in variable amounts, including dry matter, moisture content, crude protein, ether extract, ash content, and crude fiber. The percentage survival rate (PSR) in T1 was high at 87.5%, a high Feed Conversion Ratio (FCR) of 11.86, and a high Specific Growth Rate (SGR) of 37.35g. T1 had a high percentage mean weight gain (PMWG) of 566.25%, while T4 had the greatest IML (12.67 cm) of all then the diet of *C. gariepinus* without sacrificing growth performance, feed utilization, or fish health. 30% of grasshopper meal has the capacity to replace fish meal in the diet of *C. gariepinus*. Additionally, compared to imported fish meal, these indigenous protein sources are

far more affordable.

#### 4.2 Recommendation

Based on the findings of this study, it is advised that 30% grasshopper meal be added to *Clarias gariepinus* feed to promote optimal growth and lower production costs. It is advised that field experiments employing the semi-intensive system frequently used in earthen ponds be investigated as this work was conducted under experimental settings in a concrete pond.

#### References

- A.O., A.C. (1990). Official methods of Analysis (15<sup>th</sup> ed). AOAC Inc. Arlington Virginia, USA. 994P.
- Abanikannda, M.F. (2012). Nutrient digestibility and haematology of Nile tilapia (*Oreochromis niloticus*) fed with varying levels of locust (*Locusta migratoria*) meal. Bachelor of Aquaculture and Fisheries Management, Federal University of Agriculture, Abeokuta, Ogun State, July 2012.
- Abdel-Tawwab M, Ahmad MH, Khattab YA, Shalaby AM. (2010). Effect of dietary protein level, initial body weight, and their interaction on the growth, feed utilization, and physiological alterations of Nile tilapia, *Oreochromis niloticus* (L.). *Aquaculture*; **298**:267–274.
- Abdulkhaleq A, Addo S, Lawan ZA, Ebenezer A (2015) Feeding habits and conditions factor of *Oreochromis niloticus* in Lake Alau, North-eastern Nigeria. *International Journal of Fisheries and Aquatic Studies*; **3**: 406–409
- Adikwu, I. A. (2003). A Review of Aquaculture Nutrition in Aquaculture Development in Nigeria. In: Proceeding of National Workshop on Fish Feed Development and Feeding Practices in Aquaculture. Fisheries Society of Nigeria/NIFFR/FAO-NSPFS, New Bussa, Sept. 15-19: 34-42.
- Afia, O. E. and Ofor, C. O. (2016). Haematological indices of the *Clarias gariepinus* *Heterobranchius longifilis* (hybrid catfish - Heteroclarias) reared at different feeding levels. *Nigerian Journal of Agriculture, Food and Environment*; **12**(3): 6-11.
- Ahmed I. (2007). Effect of ration size on growth, body composition, and energy and protein

- maintenance requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton). *Fish Physiology Biochemical*; **33**:203–212.
- Ajenifuja, Y.F. (1998). Post-independence historical perspectives in fisheries development in Nigeria:72.
- Akinwale AO, Faturoti EO (2006). Biological performance of African Catfish (*Clarias gariepinus*) culture Recirculating system in Ibadan. *Aquaculture Engineering*; **36**:18-23.
- Akiyama, T., T. Murai, Y. Hirasawa, and T. Nose. (1984). Supplementation of various meals to fish diet for chum salmon fry. *Aquaculture*; **37**:217–222.
- Alegbeleye, W.O., S.O. Obasa, O.O. Olude, K. Otubu, and W. Jimoh. (2012). Preliminary evaluation of the nutritive value of the variegated grasshopper (*Zonocerus variegatus* L.) for African catfish *Clarias gariepinus* (Burchell. 1822) fingerlings. *Aquaculture Research*; **43**:412–420.
- Anoop, K. R., Sundar, K. S. G., Khan, B. A. and Lal, S. (2009). Common Moorhen *Gallinula chloropus* in the diet of the African catfish *Clarias gariepinus* in Keoladeo Ghana National Park, India. *Indian Birds*; **5(2)**:22-23.
- Ayinla, A.O, (2012): Challenges and investment opportunities for large-scale aquaculture farmers in Nigeria. *IIFET 2012, Tanzania Proceedings*:1-12.
- Babalola T, Apata D. 2006. Effects of dietary protein and lipid level on growth performance and body composition of African catfish *Heterobranchus longifilis* (Valenciennes, 1840) fingerlings. *Journal of Animal and Veterinary Advancement*; **5**:1073-1079.
- Balogun, A .M and Ologbobo A.D (1989): Growth performance and nutrient utilization of fingerlings of *Clarias gariepinus* (Burchell) fed raw and cooked soyabean diets. *Aquaculture*; **76**:119-126.
- Balogun, A.M and Dabrowski, K (1992) Improvement in the nutritive quality of Soy bean Meal by co-ensiling with under-utilized fish discards. *Proceedings in Fishery Society of Nigeria*; **10**:75-86
- Balogun, B.I. (2011). Growth performance and feed utilization of *Clarias gariepinus* (Teugels) fed different dietary levels of soaked *Bauhinia monandra* (Linn.) seed meal and sun-dried locust meal (*Schistocerca gregaria*). Ph.D., Dep. Biological Sciences, Faculty of Science, Ahmadu Bello University, Zaria, Nigeria. June, 2011.
- Barros, F.G., de Haro, C., Sanches-Muros, M.J., Venegas, E., Martinez Sanchez., Perez-Banon, C (2014) The potential of various insects' species for use as food for fish. *Aquaculture*: 193-201s ;422-423.
- Barroso, F.G., C. de Haro, M.-J. sanches-muros, E. venegas, A. Martinez- sanchez, and C. perez-Banon. (2014). *The potential of various insect species for use as food for fish. Aquaculture*; **422/423**:193-201.
- Bell, J.G., C. Ghioni, and J.R. Sargent. (1994). Fatty acid compositions of 10 freshwater invertebrates which are natural food organisms of Atlantic salmon parr (*Salmo salar*): *A comparison with commercial diets. Aquaculture*; **128**:301–313.
- Bondari, K., and D.C. Sheppard. (1987). Soldier fly, *Hermetia illucens* L., larvae as feed for channel catfish, *Ictalurus punctatus* (Rafinesque), and blue tilapia, *Oreochromis aureus* (Steindachner). *Aquaculture*.
- Bondari, K., and D.C. Sheppard. (1981). Soldier fly larvae as feed in commercial fish production. *Aquaculture*; **24**:103–109.
- Borthakur, S., and K. Sarma. (1998a). Protein and fat digestibility of some non-conventional fish meal replacers incorporated in the diets of fish *Clarias batrachus* (Linn.). *Environmental. Ecology*. **16**:368–371.
- Borthakur, S., and K. Sarma. (1998b). Effect of some non-conventional fish meal replacers on the growth, feed conversion and body composition of *Clarias batrachus* (Linn.) fingerlings. *Environmental. Ecology*; **16**:694–698.
- Bruton, M.N and Allanson B.R. (1980) Growth of *Clarias gariepinus* in Lake Sibaya, South Africa. *South African Journal of Zoology*; **15**: 7-15.
- Bruton, M.N. (1979a) The food and feeding behaviour of *Clarias gariepinus* (Pisces: Clariidae) in Lake Sibaya, South Africa, with emphasis on its role as a predator of cichlids. *Transactions of the Zoological Society of London*; **35**: 47-114.



- Bruton, M.N. (1979b) The breeding biology and early development of *Clarias gariepinus* (Pisces: Clariidae) in Lake Sibaya, South Africa, with a review of breeding in species of the subgenus *Clarias* (*Clarias*). *Transactions of the Zoological Society of London*; **35**: 1-45.
- Bruton, M.N. (1979c) The survival of habitat desiccation by air-breathing Clariid catfishes. *Environmental Biology of Fishes*; **4**: 273-280.
- Bulet P, Hetru C, Dimarcq JL, Hoffmann D (1999) Antimicrobial peptides in insects; structure and function. *Developmental and Comparative Immunology*; **23**: 329–344.
- Carolsfeld J, Harvey B, Ross C, Baer A (eds) (2003) *Migratory fishes of South America. Biology, Fisheries and Conservation Status*. World Fisheries Trust. *The World Bank International Development Research Centre, Victoria, BC*.
- Chavez-Sanchez, M.C., C.A. Martinez-Palacios, G. Martinez-Perez, and L.G. Ross. (2000). Phosphorus and calcium requirements in the diet of the American cichlid *Cichlasoma urophthalmus* (Günther). *Aquaculture. Nutrition*; **6**: 1–9.
- Chernysh S, Gordya N, Suborova T (2015) Insect antimicrobial peptide complexes prevent resistance development in bacteria. *PLoS One*; **10**: e0130788.
- Chika, I.K and Nathaniel, O. (2014). The culture of Catfish in Nigeria: Progress, prospects and problems. *Advances in Aquaculture and Fisheries Management*; **2(2)**: 116-119.
- Chukwuma, O. and Henry, U. (2012). Effect of water hardness on fertilization and hatchery success of *Clarias gariepinus* (Burchell, 1822) and *Heterobranchus longifilis* (Valenciennes, 1840) eggs fertilized with *Clarias gariepinus* sperm. *Asian Fisheries Science*; **25**: 270-277
- Cranney, A., Horsley, T. Donnell, S., Weiler, H., Puil, L., Ooi, D., Atkinson, S., Ward, L., Moher, D., Hanley, D., Fang, M., Yazdi, Z., Garritty, C., Sampson, C., Barrowman, C., Tsertsvadze, A. & Mamaladze, V. (2007). *Effectiveness and safety of vitamin D in relation to bone health. Evidence Report Technology Assess*; **58**: 1–235.
- Darnell RM (1958) Food habits of fishes and larger invertebrates of Lake Pontchartrain, Louisiana, an Estuarine Community. *Institute of Marine Science*; **5**: 353–416.
- De Moor, I and Bruton, M.N. (1988). Atlas of alien and translocated indigenous aquatic animals in southern Africa. South African National Scientific Programmes Report No. 144. CSIR.
- Dhirendra, K., (2002). Physico-chemical characteristics of two ecologically different ponds. The fifth Indian Fisheries Forum Proceedings. (Eds. Ayyappan, S., Jena, J.K. and Mohan, M.) AFS13 and AOA, Bhubaneswar, India: 193-198.
- Dyer DC, Perissinotto R, Carrasco NK (2015) Temporal and spatial dietary dynamics of the longspine glassy (Ambassis ambassis) in the St. Lucia estuarine system, iSimangaliso Wetland Park. *Water SA*; **41**: 91–104.
- Ebenso, I.E and M.T. Udo. (2003). Effect of live maggot on growth of the Nile perch, *Oreochromis niloticus* (Cichlidae) in South Eastern Nigeria. *Global Journal of Agricultural Science*; **2**: 72–73
- Effiong, B. N., Sanni, A and Fakunle, J. O. (2009). Effect of partial replacement of fish meal with duckweed (*Lemna pauciscoctata*) meal on the growth performance of *Heterobranchus longifilis* fingerlings. *Report and Opinions*, 1(3), 76-81.
- Ezenwa, B, (1994). Aquaculture development and research in Nigeria: In aquaculture development and research in Sub-Saharan Africa, National Reviews. CIFA Technical paper, 223 supplement. FAO, Rome. Pp 45-79.
- Fagbenro, O. A., and Davies S.J., (2003). Use of high percentages of soya protein concentrate as fishmeal substitute in practical diets for African catfish growth, feed utilization and digestibility. *Journal of Aquaculture* 16(1).
- Fagbenro, O. A., Smith, M., & Amoo, A. I. (2012). (Digitaria exilisstapf) Meal Compared with Maize and Sorghum Meals as Dietary Carbohydrate Source for Nile Tilapia (*Oreochromis niloticus*). *Israel Journal of Aquaculture*, 2(52), 3-1.
- FAO (2005-2012). *Cultured Aquatic Species*

- Information Programme. *Clarias gariepinus*. Cultured Aquatic Species Information Programme. Text by Rakocy, J. E. In: FAO Fisheries and Aquaculture Department [online]. Rome. Updated 18 February 2005. [Cited 11 September 2012].
- FAO (2010). The state of world fisheries and aquaculture. FAO, Fisheries and Aquaculture Development, Rome, Italy.
- FAO. (2014). The state of world fisheries and aquaculture- Opportunities and challenges. Rome: Food and Agriculture organization of the United Nations.
- Fasakin, E.A., A.M. Balogun, and O.O. Ajayi. (2003). Evaluation of full-fat and defatted maggot meals in the feeding of clariid catfish *Clarias gariepinus* fingerlings. *Aquaculture*. Reserved. 34:733–738.
- Ferrareze M, Nogueira MG, Casatti L (2015) Differences in ichthyofauna feeding habits among lateral lagoons and the river channel in a large reservoir. *Brazilian Journal of Biology* 75: 380–390
- Finke, M.D. (2007). Estimate of chitin in raw whole insects. *Zoology Biology*. 26:105–115.
- Finke, MD. (2015). Complete nutrient content of four species of commercially available feeder insects fed enhanced diets during growth. *Zoo Biology*, 34(6), 554-564.
- Francis, G., Makkar, H. P. S and Becker, K. (2001) Anti nutritional factors present in plant derived alternative fish feed ingredients and their effects in fish. *Aquaculture* 199: 197-227.
- Gabriel, U.U., Akinrotimi, O.A. D. O. Bekibele2, D. N Onunkwo2 and P. E. Anyanwu (2007). Locally produced fish feed: potentials for aquaculture development in subsaharan Africa. *African Journal of Agricultural Research*, 2(7), pp. 287-295.
- Gasco, L., F. Gai, G. Piccolo, L. Rotolo, C. Lussiana, P. Molla, and S. Chatzifotis. (2014b). Substitution of fish meal by *Tenebrio molitor* meal in the diet of *Dicentrarchus labrax* juveniles. In: *Insects to Feed the World*, The Netherlands, 14–17 May 2014. p. 80.
- Gasco, L., M. Belforti, L. Rotolo, C. Lussiana, G. Parisi, G. Terova, A. Roncarati, and F. Gai. (2014a). Mealworm (*Tenebrio molitor*) as a potential ingredient in practical diets for rainbow trout (*Oncorhynchus mykiss*). In: *Insects to Feed the World*, The Netherlands, 14–17 May 2014. p. 78.
- Gbadamoal O. K., Daramola, J. A, and Osungbemi (2007). Growth performance and nutritional utilization of vitamin C in diet of African catfish fingerlings. *Acta Zoological science* 58:763-766.
- Ghosh, A. K. & Joshi, S. R. (2008). Disorders of calcium, phosphorus and magnesium metabolism. *Journal of Association of Physiology*, 56, 613–21.
- Halver, J.E and Hardy, R.W(2002) Fish Nutrition. In Sargent, J.R., Tocher, D.R and Bell,G., *The lipids*, 3<sup>rd</sup> Edition, Academic Press, Carlifornia, 182-246
- Hecht, T. and S. Appelbaum, (1987). Notes on the growth of Israeli sharptooth catfish (*Clarias gariepinus*) during the primary nursing phase. *Aquaculture*, 63: 195–204.
- Hecht, T., Uys, W and Britz, P.J. (Editors). (1988). *Culture of sharptooth catfish, Clarias gariepinus*, in southern Africa. National Scientific Programmes Unit: CSIR, SANSP Report 153, 1988, pp 146.
- Henry M, Gasco L, Piccolo G, Fountoulaki E. (2015). Review on the use of insects in the diet of farmed fish: past and future. *Anim Feed Sci Tech*. 203:1-22.
- Holick, M. F. (2008). Diabetes and the Vitamin D connection. *Current Diabetes Report*, 8, 393–398.
- Hossain, M.A., M.N. Islam, and M.A. Alim. (1993). Evaluation of silkworm pupae meal as dietary protein source for catfish (*Heteropneustes fossilis* Bloch). In: *Fish nutrition in Practice: 4th International Symposium on Fish Nutrition and Feeding*. Biarritz, France, 24–27 June 1991. p.785–791.
- Hossain, M.A., N. Nahar, and M. Kamal. (1997). Nutrient digestibility coefficients of some plant and animal proteins for rohu (*Labeo rohita*). *Aquaculture* 151:37–45.
- Hossain, M.A., N. Nahar, M. Kamal, and M.N. Islam. (1992). Nutrient digestibility coefficients of some plant and animal proteins for tilapia (*Oreochromis mossambicus*). *J. Aquacult. Trop*. 7:257–265.

- Igah, S. A. (2008). Target food sources for formulating complementary/Supplementary foods for catfish purposes - a review. *Bayero Journal of Pure and Applied Science*, 2, 11-23.
- IGFA, (2001). Database of IGFA angling records until 2001. IGFA, Fort Lauderdale, USA.
- Johnels, A.G. 1957. The mode of terrestrial locomotion in *Clarias*. *Oikos* 8: 122-129.
- Jubb, R.A. 1967. Freshwater fishes of southern Africa. A.A. Balkema, Cape Town; viii + 248 pp.
- Inyang. E. B., (2001). Factors affecting aquaculture production in Akwa Ibom State. Unpublished M.Sc project report to the department agricultural extension and rural development, University of Ibadan, Ibadan. 80 pp. Maxiprints, Awka, Nigeria.
- Ji, W.X., Y. Wang, and J.Y. Tang. (2010). Apparent digestibility coefficients of selected feed ingredients for Japanese sea bass (*Lateolabrax japonicus*) reared in sea water. *J. Fish. China*. 34:101–107
- Johri, R., R. Singh, and P. K. Johri. (2011a). Studies on ovarian activity in formulated feed treated *Clarias batrachus* Linn. *J. Exp. Zoology*, India. 14:111–115.
- Johri, R., R. Singh, and P. K. Johri. (2011b). Histopathological examination of the gill, liver, kidney, stomach, intestine, testis and ovary of *Clarias batrachus* Linn. during the feeding on different formulated feeds. *J. Exp. Zoology*, India. 14:77–79.
- Johri, R., R. Singh, and P.K. Johri. (2010). Effect of different formulated plant and animal diet on hematology of *Clarias batrachus* Linn. under laboratory conditions. *Biochem Cell Arch*. 10:283–291.
- Jozefiak A, Engberg RM (2017) Insect proteins as a potential source of antimicrobial peptides in livestock production. A review. *Journal of Animal and Feed Sciences* 26: 1–12
- Kadye, T. (2011). Assessing the impacts of invasive non-native African sharptooth catfish *Clarias gariepinus*. PhD thesis, Rhodes University.
- Keat-Chuan, N. L., Aun-Chuan, O. P. L., Wong, W. L. L. & Khoo, G. (2017). A Review of Fish Taxonomy Conventions and Species Identification Techniques. *Journal of Survey in Fisheries Sciences*, 4(1), 54-93.
- Kourimska L, Adamkova A (2016) Nutritional and sensory quality of edible insects. *NFS Journal* 4: 22
- Kroeckel, S., A.G.E. Harjes, I. Roth, H. Katz, S. Wuertz, A. Susenbeth, and C. Schulz. (2012). When a turbot catches a fly: Evaluation of a pre-pupae meal of the black soldier fly (*Hermetia illucens*) as fish meal substitute-growth performance and chitin degradation in juvenile turbot (*Psetta maxima*) *Aquaculture* 364/365:345–352.
- Kumar, V., A.K. Singh, H.P. Makkar, G. De Boeck, and K. Becker. (2012). Phytate and phytase in fish nutrition. *J. Anim. Physiology. Animal. Nutrition*. 96:335–364.
- Ladon, S. (2000). A fish farmer's guide to Understanding water Quality, department of Animal Sciences Illinois, Indiana Sea, Grant programme, purdue University.
- Laurin, M. & Reisz, R. R. (1995). Biology of Actinopterygii. *Zoological Journal of the Linnean Society*, 113 (2), 165–223.
- Lindsay, G.J.H., M.J. Walton, J.W. Adron, T.C. Fletcher, C.Y. Cho, and C.B. Cowey. (1984). The growth of rainbow trout (*Salmo gairdneri*) given diets containing chitin and its relationship to chitinolytic enzymes and chitin digestibility. *Aquaculture* 38:315–334
- Lock ER, Arsiwalla T, Waagbø R (2016) Insect larvae meal as an alternative source of nutrients in the diet of Atlantic salmon (*Salmo salar*) postsmolt. *Aquaculture Nutrition*; 22: 1202–1213.
- Lock, E.J., T. Arsiwalla, and R. Waagbø. (2014). Insect meal: A promising source of nutrients in the diet of Atlantic salmon (*Salmo salar*). In: *Insects to Feed the World*, The Netherlands, 14–17 May 2014:74.
- Madhuri, S., Shrivastav, A. B., Sahni, Y. P. and Govind, P. (2012). Overview of the treatment and control of common fish diseases. *Internal Research Journal of Pharmacy*; 3(4): 2230-8407.
- Madu, C.T., and E.B.C. Ufodike. (2003). Growth and survival of catfish (*Clarias anguillaris*) juveniles fed live tilapia and maggot as unconventional diets. *Journal of Aquacultural Science*; 18:47–51
- Madu, C.T., Sogbesan, O.A. and Ibiyo, L.M.O. (2003). Some non-conventional fish feed resources in Nigeria. In: A.A. Eyo (Ed.),

- Proceeding joint FISON/NIFFR/SPFS National workshop on fish feed and Feeding practices in Aquaculture held, National Institute for Freshwater Fisheries Research, 15<sup>th</sup>–19<sup>th</sup> September, 2003. New-Bussa: 73–82.
- Magalhes R, Sanchez-Lopez A, Silva Leal R, Martinez-Llorens S, Oliva-Teles A, Peres H (2017) Black soldier fly (*Hermetia illucens*) pre-pupae meal as a fish meal replacement in diets for European seabass (*Dicentrarchus labrax*). *Aquaculture*; **476**: 79–85.
- Makkar HP, Tran G, Heuze V, Ankers P. 2014. State-of-the-art on use of insects as animal feed. *Animal Feed Science Technology*; **197**: 1–33.
- Malde, M. K., Bugel, S., Kristensen, M., Malde, K., Graff, I. E. & Pedersen., J. I. (2010). Calcium from salmon and cod bone is well absorbed in young healthy men: A double-blinded randomised crossover design. *Nutritional Metabolism*; **2(3)**: 7–61.
- Manzano-Agugliaro, F., M.J. Sanchez-Muros, F.G. Barroso, A. Martínez-Sánchez, S. Rojo, and C. Pérez-Bañón. (2012). *Insects for biodiesel production*. *Renew. Sustain. Energy Review*; **16**: 3744–3753.
- Matsumiya M, Arakane Y, Haga A, Muthukrishnan S, Kramer KJ (2006) Substrate specificity of chitinases from two species of fish, greenling, *Hexagrammos otakii*, and common mackerel, *Scomber japonicus*, and the insect, tobacco hornworm, *Manduca sexta*. *Bioscience, Biotechnology and Biochemistry*; **70**: 971–979.
- Médale, F., R. Le Boucher, M. Dupont-Nivet, E. Quillet, J. Aubin, and S. Panserat. (2013). Des aliments à base de végétaux pour les poissons d'élevage. INRA Production. *Animal*; **26**: 303–316.
- Merron, G.S. (1993). Pack-hunting in two species of catfish, *Clarias gariepinus* and *C. ngamensis*, in the Okavango Delta, Botswana. *Journal of Fish Biology*; **43**: 575–584.
- Momoh, P. and Solomon, R. J. (2017). Physiochemical parameters of Catfish (*Clarias gariepinus*) fed a combination of rice bran and Irish potatoes. *Direct Research Journal of Veterinary Medical Animal Science*; **2(2)**: 36–50.
- Nandeesh, M.C., B. Gangadhara, T.J. Varghese, and P. Keshavanath. (2000). Growth response and flesh quality of common carp, *Cyprinus carpio* fed with high levels of non-defatted silkworm pupae. *Asian Fish. Science*; **13**: 235–242.
- Nandeesh, M.C., G.K. Srikantha, P. Keshavanatha, T.J. Varghese, N. Basavarajaa, and S.K. Dasa. (1990). Effects of non-defatted silkworm-pupae in diets on the growth of common carp, *Cyprinus carpio*. *Biology. Wastes*; **33**: 17–23.
- Narayanan S, Modak JK, Ryan CS, Garcia-Bustos J, Davies JK, Roujeinikova A (2014) Mechanism of Escherichia coli resistance to pyrrolic acid. *Antimicrobial Agents Chemotherapy*; **58**: 2754–2762.
- Newton, L., C. Sheppard, D.W. Watson, G. Burtle, and R. Dove. (2005). Using the black soldier fly, *Hermetia illucens*, as a value-added tool for the management of swine manure. Report for Mike Williams, Director of the Animal and Poultry Waste Management Center, North Carolina State University. [http://www.cals.ncsu.edu/waste\\_mgt/smithfield\\_projects/phase2report05/cd\\_web%20files/A2.pdf](http://www.cals.ncsu.edu/waste_mgt/smithfield_projects/phase2report05/cd_web%20files/A2.pdf). (Verified 3 Feb. 2015.)
- Ng, W.K., F.L. Liew, L.P. Ang, and K.W. Wong. (2001). Potential of mealworm (*Tenebrio molitor*) as an alternative protein source in practical diets for African catfish, *Clarias gariepinus*. *Aquaculture*; **32(1)**: 273–280
- Njidda, A.A and C.E Isidahomen (2010): Haematology, blood chemistry and carcass characteristics of growing rabbits Ibukun Grace Olaleye- Effects of Grasshopper meal in the Diet of *Clarias gariepinus* Fingerlings fed grasshopper meal as a substitute for fishmeal. *Pakistan Veterinary Journal*; **30(1)**: 7–12.
- Nunes AJP, Sa MVC, Browdy CL, Vazquez-Anon M (2014) Practical supplementation of shrimp and fish feeds with crystalline amino acids. *Aquaculture*; **431**: 20–21
- Odum, H.T. (1959). Analysis of Diurnal Oxygen curves for the Essays of Marine Bays. In: proceeding of the first international conference on waste Disposal in the marine environment. Retrieved from <https://doi.org/10.2307/1929948>.

- Ogunji, J.O., J. Nimptsch, C. Wiegand, and C. Schulz. (2007). Evaluation of the influence of housefly maggot meal (mameal) diets on catalase, glutathione S-transferase and glycogen concentration in the liver of *Oreochromis niloticus* fingerling. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*; **147(4)**: 942-947.
- Ogunji, J.O., W. Kloas, M. Wirth, C. Schulz, and B. Rennert. (2008a). Housefly maggot meal (mameal) as a protein source for *Oreochromis niloticus* (Linn.). *Asian Fish. Science*; **21**:319–331.
- Ogunji, J.O., W. Kloas, M. Wirth, N. Neumann, and C. Pietsch. (2008b). Effect of housefly maggot meal (mameal) diets on the performance, concentration of plasma glucose, cortisol and blood characteristics of *Oreochromis niloticus* fingerlings. *Journal Animal Physiology. Animal. Nutrition*; **92**:511–518
- Okoye, F.C and Nnaji, J.C (2004) Effect of substituting fishmeal with grasshopper meal on the growth and food utilization of the Nile Tilapia, *Oreochromis niloticus* fingerlings.
- Oliveira, V., Poleto, S.L and Venere, P.C (2005) Feeding of juvenile pirarucu (*Arapaima gigas*, *arapaimidae*) in their natural environment, lago Quatro Bocas, Araguaina-MT, Brazil. *Neotropical Ichthyology*; **3**: 312–31
- Olurin KB, Olojo AA, Olukoya OA. (2006) Growth of African catfish *Clarias gariepinus* fingerlings fed different levels of cassava. *World Journal of Zoology*; **1(1)**:54-56.
- Piccolo, G., S. Marono, L. Gasco, F. Iannaccone, F. Bovera, and A. Nizza. (2014). Use of *Tenebrio molitor* larvae meal in diets for gilthead sea bream *Sparus aurata* juveniles. In: *Insects to Feed the World*, The Netherlands, 14–17 May 2014:76.
- Picker, M. and Griffiths, C. (2011). *Alien & invasive animals, a South African perspective*. Struik Publishers, Cape Town.
- Rahman, M.A., M. Zaher, M.A. Mazid, M.Z. Haque, and S.C. Mahata. (1996). Replacement of costly fish meal by silkworm pupae in diet of mirror carp (*Cyprinus carpio* L.). *Pakistani Journal of Science*; **39**:64–67.
- Ramos-Elorduy J (1998) *Creepy Crawly Cuisine: The Gourmet Guide to Edible Insects*. Park Street Press, Paris, France.
- Rangacharyulu, P.V., S.S. Giri, B.N. Paul, K.P. Yashoda, R.J. Rao, N.S. Mahendrakar, S.N. Mohanty, and P.K. Mukhopadhyay. (2003). Utilization of fermented silkworm pupae silage in feed for carps. *Bioresources Technology*; **86**:29–32
- Rangaswamy CP (2006) Physiology of digestion in fish and shrimp. In: Ali SA (ed.) *Training Manual on Shrimp and Fish Nutrition and Feed Management: 2–9*. Central Institute of Brackish Water Aquaculture, Chennai
- Reed D, Knaapila A (2010) Genetics of taste and smell: poison and pleasure. *Progress in Molecular Biology and Translational Science*; **94**: 213–240.
- Ribeiro AR, Biagioni RC, Smith WS (2014) Estudo da dieta natural da ictiofauna de um reservatorio centenário, São Paulo, Brasil. *Iheringia, Series Zoological, Porto Alegre*; **104**: 404–412.
- Ricardo B. R., Edward, O. W., Gloria, A., Nicolas, B., Masaki, M., Guillaume, L and Guillermo, O. (2017). Phylogenetic classification of bony fishes. *BMC Evolutionary Biology*; **17(1)**: 162-176.
- Robins, C.R., Bailey, R.M., Bond, C.E., Brooker, J.R., Lachner, E.A., Lea, R.N. & Scott, W.B. (1991). World fishes important to North Americans. Exclusive of species from the continental waters of the United States and Canada. *American Fish Society. Spec. Published*; **21**: 243.
- Robinson, E. H., Li, M. H. & Manning, B. B. (2001). A practical guide to nutrition, feeds and Feeding of Catfish. *Mississippi Agricultural and Forestry Experiment Station Bulletin* No. 1113 (Second Revision); **1(1)**: 20–27.
- Robinson, E. H., Weirich, C. R. and Li, M. H. (1994). Feeding Catfish. *Mississippi Agricultural and Forestry Experiment Station Bulletin* No. 1019.
- Rudkowska, I. B., Marcotte, G., Pilon, C., Lavigne, A. & Vohl, M. C. (2010). Fish nutrients decrease expression levels of tumor necrosis factor- $\alpha$  in cultured human macrophages. *Journal of Physiological Genomics*; **40(8)**: 189–194.
- Safriel, O. and Bruton, M.N. (1984). A

- cooperative aquaculture research programme for South Africa. South African National Scientific Programmes Report 89. CSIR, Pretoria:79.
- Sanchez-Muros M-J, Barroso FG, Manzano-Agugliaro F. 2014. Insect meal as renewable source of food for animal feeding: a review. *Journal of Clean Production*; **65**:16-27.
- Sealey, W.M., T.G. Gaylord, F.T. Barrows, J.K. Tomberlin, M.A. McGuire, C. Ross, and S. St-Hilaire. (2011). Sensory analysis of rainbow trout, *Oncorhynchus mykiss*, fed enriched black soldier fly prepupae, *Hermetia illucens*. *Journal. World Aquaculture Society*; **42**:34–45.
- Shyama, S., and P. Keshavanath. (1993). Growth response of Tor khudree to silkworm pupa incorporated diets. In: Fish Nutrition in Practice: 4th International Symposium on Fish Nutrition and Feeding. Biarritz, France, 24–27 June 1991: 779–783.
- Silva-Camacho DS, Santos JNS, Gomes RS, Araujo FG (2014) Eco morphological relationships among four characiformes fish species in a tropical reservoir in South-eastern Brazil. *Zoologia*; **31**: 28–34.
- Skelton, P. (2001). A complete guide to the freshwater fishes of Southern Africa. Struik Publishers, Cape Town.
- Sogbesan A, Ugwumba A. (2008). Nutritional evaluation of termite (*Macrotermes subhyalinus*) meal as animal protein supplements in the diet of *Heterobranchus longifilis* (Valenciennes, 1840) fingerlings. *Turkish Journal of Fishery and Aquaculture Science*; **8**:149-157.
- Sogbesan OA. Culture and utilization of some nonconventional animal protein feedstuffs in the diet of *Heterobranchus longifilis* (Valenciennes, 1840). Thesis PhD. Department of Zoology, Faculty of Science, University of Ibadan, Nigeria, 2007: 252.
- Sogbesan, A, and Ugwumba, A. (2008). Nutritional evaluation of termite (*Macrotermes subhyalinus*) meal as animal protein supplements in the diets of *Heterobranchus longifilis* (Valenciennes, 1840) fingerlings. *Turkish Journal of Fisheries and Aquatic Sciences*; **8**:149–157.
- Sogbesan, A.O., N. Ajuonu, B.O. Musa, and A.M. Adewole. (2006). Harvesting techniques and evaluation of maggot meal as animal dietary protein source for *Heteroclaris* in outdoor concrete tanks. *World Journal of Agricultural Science*; **2**:394–402
- Spataru P, Hephher B, Halevy A (1980) The effect of the method of supplementary feed application on the feeding habits of carp (*Cyprinus carpio* L.) with regard to natural food in ponds. *Hydrobiologia*; **72**: 171–178.
- St-Hilaire, S., C. Sheppard, J.K. Tomberlin, S. Irving, L. Newton, M.A. McGuire, E.E. Mosley, R.W. Hardy, and W. Sealey, W. (2007a). Fly prepupae as a feedstuff for rainbow trout, *Oncorhynchus mykiss*. *Journal of World Aquaculture Society*; **38**:59–67.
- Swamy, H.V.V., and K.V. Devaraj. (1994). Nutrient utilization by common carp (*Cyprinus carpio* Linn) fed protein from leaf meal and silkworm pupae meal-based diets. *Indian Journal of Animal Nutrition*; **11**:67–71.
- Tacon, A. G. J. & Metian, M. (2013). Fish matters, importance of aquatic foods in human nutrition and global food supply. *Review Fisheries Science*; **21(5)**:22–38.
- Tamanna, N., Mahmood, N. (2015) Food processing and Maillard reaction products: effect on human health and nutrition. *International Journal of Food Science*; **2015**: 526762.
- Ulrich SP (2017) Native fish species boosting Brazilian's aquaculture development. *Acta Fisheries and Aquatic Resources*; **5**:1-9.
- Van Hius, A., Itterbeeck, J. V., Klunder, H., Mertens, E., Halloran, A., Muir, G, and Vantomme, P. (2013). Edible Insects-Future prospects for food and feed security. *FAO Forestry Paper*; 171.
- Viveen, W.J.A.R., Richter, C.J.J., Van Jassen, J.A.L. and Huisman, E.A. (1977). Practical Manual for the culture of the African catfish (*Clarias gariepinus*). Joint publication of Directorate General International Cooperation of the Ministry of Foreign Affairs. The Hague, The Netherlands, Department of Fish Culture and Fisheries of the Agricultural University of Wageningen, the Netherlands and Research Group for Comparative Endocrinology, Department of Zoology of the University of Utrecht, the Netherlands: 94.

- Willoughby, N.G. & Tweddle, D. 1978. The ecology of the catfish *Clarias gariepinus* and *Clarias ngamensis* in the Shire Valley, Malawi. *Journal of Zoology*; **186**: 507-534.
- World Health Organization (WHO) (2013). Water-related diseases. (Available at [www.who.int/water\\_sanitation\\_health/diseases/malnutrition/en/](http://www.who.int/water_sanitation_health/diseases/malnutrition/en/)). Accessed February 2017.
- Yi HY, Chowdhury M, Huang YD, Yu XQ (2014) Insect antimicrobial peptides and their applications. *Applied Microbiology and Biotechnology*; **98**: 5807–5822.
- Yovita, J. M. (2007). The Effects of Dissolved Oxygen on Fish Growth in Aquaculture. (An MSc. Thesis, The United Nations University.120, Reykjavik, Iceland:30.
- Zhang, J.B., L.Y. Zheng, P. Jin, D.N. Zhang, and Z.N. Yu. (2014). Fishmeal substituted by production of chicken manure conversion with microorganisms and black soldier fly.

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